

# OliCheck v1.0.5 - User Guide

## Introduction

OliCheck is a program designed to test the validity of potential microarray probes by considering the possibility of cross-hybridization with non-target coding sequences. This analysis can be performed within a single genome or between several different strains/organisms. For more information on the algorithm, see Charbonnier *et al.*, unpublished.

For specific problems/questions, see the Notes section located at the end of this document or send an email to [yvan.charbonnier@genomic.ch](mailto:yvan.charbonnier@genomic.ch).

## Installing OliCheck

Before you install OliCheck, make sure that your computer system meets the minimum requirements for that product. See System Requirement (page 5) for more information.

### ***To install OliCheck:***

Run: setup.exe

Follow the installation instructions displayed on the screen. You can perform a Compact, Typical, or Custom installation. The Compact installation includes only the files you need to run OliCheck (i.e. the software and its manual). The Typical installation includes all the files you need to run OliCheck plus the BLAST executable and example files. Choosing the Custom installation, you can install any combination of the following:

- OliCheck Application

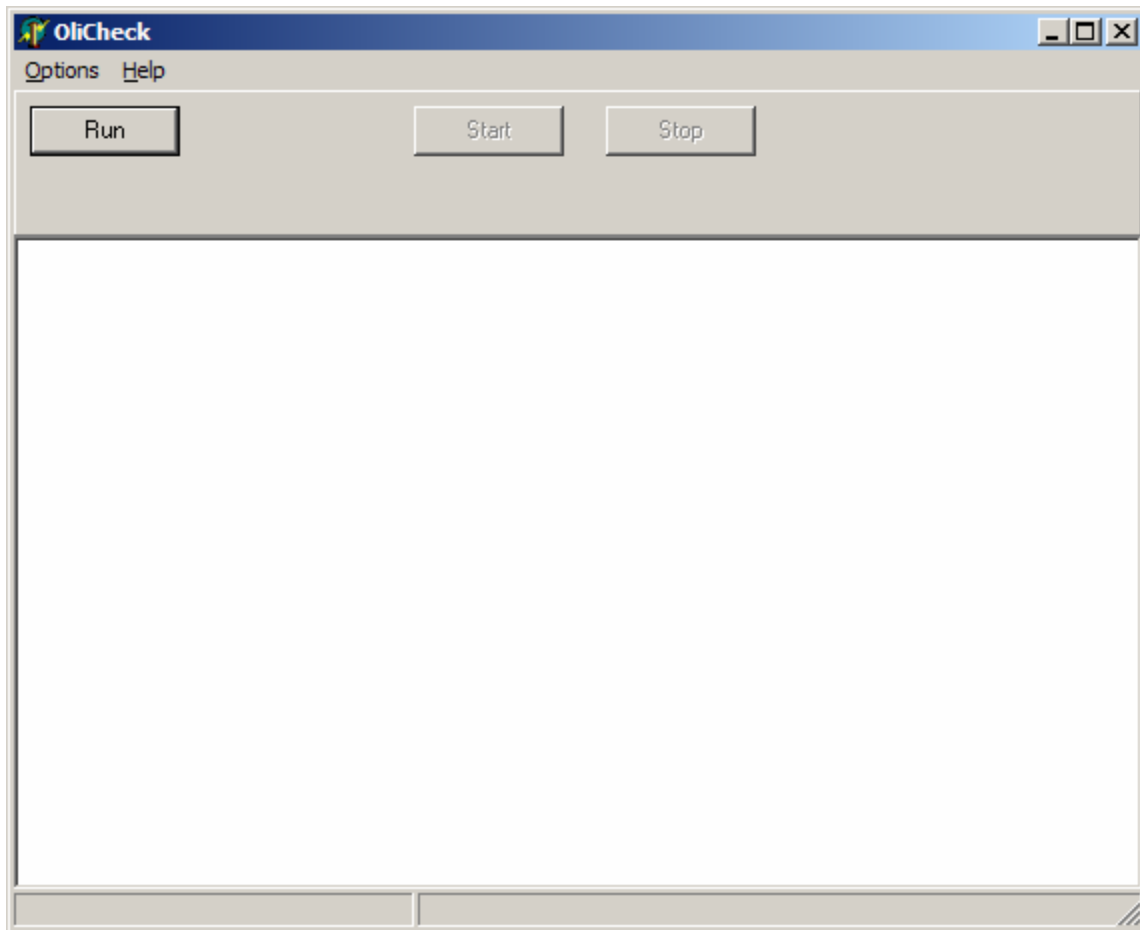
- BLAST Executable

- OliCheck Example Files:

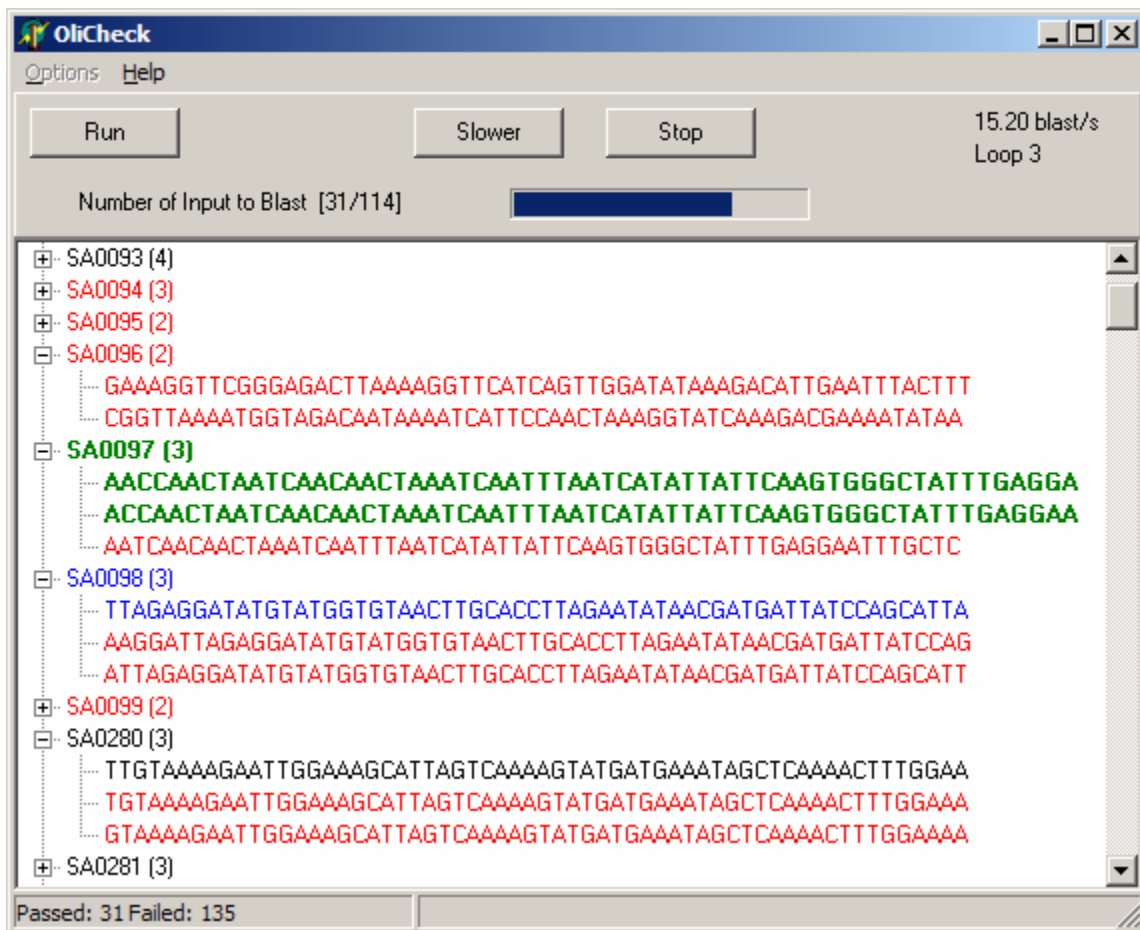
- BLAST Example Database
- Array Designer™™ Example File

## Using OliCheck

OliCheck requires one input file, as described on page 7. This file must be in the format used by Array Designer™, as its standard output (see page 8). All outputs generated by OliCheck will also be provided in this format. The specifications of a run of OliCheck are defined by clicking on the Options menu located in the upper left-hand corner of the program window (Figure below). A detailed review of these parameters can be found on page 3.

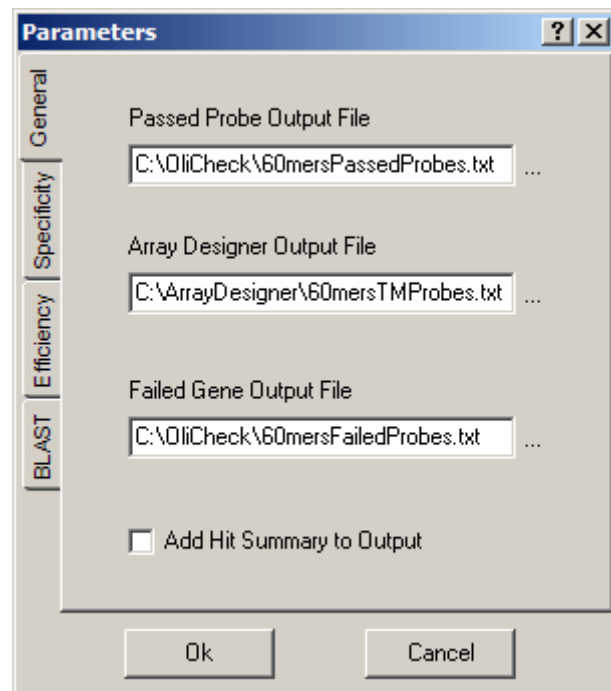


After having completed the setup, click 'OK' on the Parameters box. Then, select 'Run' in the main window. OliCheck will load the desired Array Designer™ file and test the probes according to the selected parameters. The main window provides the user with a variety of information on the status of the process. It displays the current position in the BLAST queue, the number of BLASTs performed per second, the loop number, the number of genes that have probes that passed/failed and the gene names and probe sequences themselves. A gene name/probe sequence is color-coded in the right-hand window depending on whether it has passed (green) or failed (red) the OliCheck process. When a probe is in the process of being BLASTed, it is highlighted in blue. The Typical installation allows the user to run an example illustrating that process.



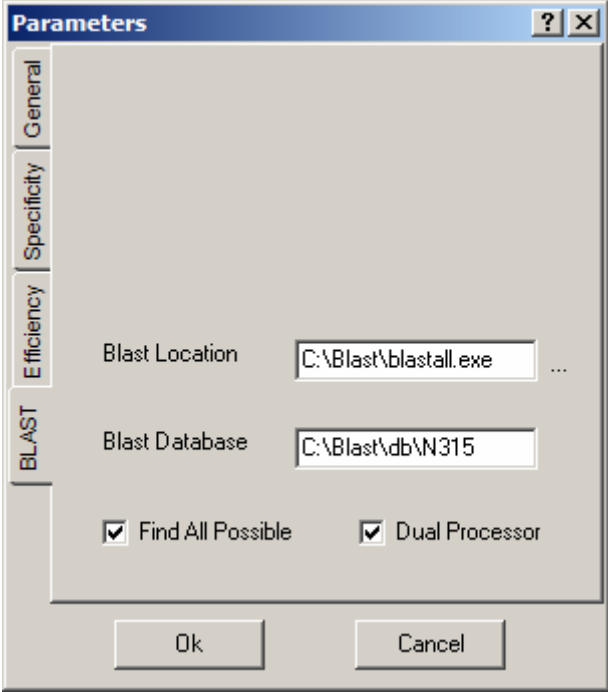
## OliCheck Parameters

The first step in initiating an OliCheck test is to select the files using the **General submenu**. The files that OliCheck creates will follow standard formatting conventions. The “passed probe” file will be returned in Array Designer™ format. It will contain the thermodynamics parameters and comments taken from the Array Designer™ input file, to describe each probe that OliCheck validated as being appropriate. The “failed gene” file will contain a simple list of all genes for which no acceptable probe has been found. If the *Add Hit Summary to Output* is checked, the “passed probe” file will contain an extra column with the



name of the homologous gene(s) that the probe can detect. This will be useful only for checking potential probes against another genome than the one used to generate the probe set.

Using the **BLAST submenu**, the user can select the BLAST database to be used to validate the probe set. The name of the database, as defined by one's local installation of BLAST, should be typed here along with the location of your blast executable. The *Find All Possible* box instructs OliCheck not to stop after having found a probe for a given gene that satisfies user-defined parameters. The *Dual Processor* allows OliCheck taking advantage of BLAST's ability to split its processes on two processors.

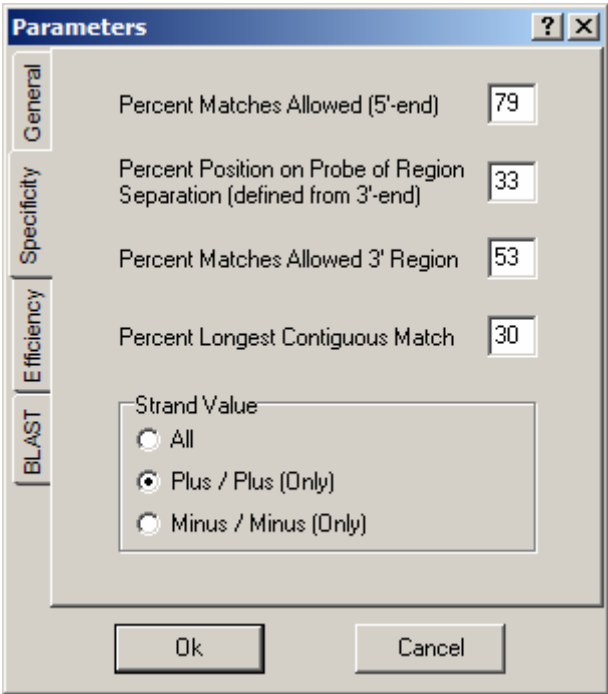


The image shows a 'Parameters' dialog box with the 'BLAST' tab selected. The 'General' tab is also visible. The 'BLAST' tab contains the following fields and options:

- Blast Location:** C:\Blast\blastall.exe
- Blast Database:** C:\Blast\db\N315
- ☒ Find All Possible
- ☒ Dual Processor

Buttons at the bottom: Ok, Cancel.

The **Specificity submenu** allows defining values for the parameters used in the Specificity test (see Charbonnier *et al.* for further description). The values listed in the adjacent figure are the defaults used by OliCheck. These parameters are a mathematical interpretation of the experimentally defined specificity parameters in Hughes *et al.* (Nat Biotech, 2001; 19(4):342-7). The results from the paper refer to a number of matches/mismatches for 60-mer oligonucleotides. The value used in OliCheck is defined as a percentage to allow dealing with different oligonucleotide lengths. The first parameter indicates the percentage of matches allowed in the solution-end of the probe (5'-end). The second parameter



The image shows a 'Parameters' dialog box with the 'Specificity' tab selected. The 'General' tab is also visible. The 'Specificity' tab contains the following fields and options:

- Percent Matches Allowed (5'-end):** 79
- Percent Position on Probe of Region Separation (defined from 3'-end):** 33
- Percent Matches Allowed 3' Region:** 53
- Percent Longest Contiguous Match:** 30
- Strand Value:**
  - ☐ All
  - ☒ Plus / Plus (Only)
  - ☐ Minus / Minus (Only)

Buttons at the bottom: Ok, Cancel.

allows specifying the portion of the probe that will be considered as the 5'-end and 3'-end regions. The third parameter indicates the percentage of matches allowed in the surface-end of the probe (3'-end). A last parameter defines the longest contiguous match allowed. Finally, OliCheck is able to restrict its filtering to the Plus, Minus or Both strands of the BLAST alignment.

The **Efficiency submenu** allows defining values for parameters used in the Efficiency test (see Charbonnier *et al*). Values listed in the adjacent figure are the defaults used by OliCheck. The first parameter specifies the total number of mismatches allowed in the whole BLAST alignment between the probe and its target. The second parameter allows specifying the portion of the probe that will be considered as the 5'-end and 3'-end regions. The third parameter sets the maximum percentage of mismatches allowed in the solution-end of the probe.

The screenshot shows the 'Parameters' dialog box with the 'Efficiency' tab selected. The dialog has four tabs: 'General', 'Specificity', 'Efficiency', and 'BLAST'. The 'Efficiency' tab contains the following settings:

- Percent Total Mismatches Allowed: 67
- Percent Position on Probe of the Last Allowed Mismatch (3' end): 50
- Percent Mismatches Allowed in this Region: 0
- Strand Value:
  - ☐ All
  - ☒ Plus / Plus (Only)
  - ☐ Minus / Minus (Only)

At the bottom are 'Ok' and 'Cancel' buttons.

## Notes

At present, OliCheck contains a memory leak that might cause problems if it is sequentially used to analyze large Array Designer™ files. To avoid this problem, simply quit OliCheck between runs: the program will free all memory associated with it on shut-down thereby eliminating any possible memory leak.

With Microsoft Windows™ XP using graphic themes, user may experience some graphical rendering problems with the vertical tabs. The graphical interface is fully rendered with the Windows™ Classic Theme.

## System Requirement

OliCheck requires additional software:

NCBI Blast provided in the installation executable or freely downloadable on the NCBI ftp server (<ftp.ncbi.nih.gov/blast/executables/LASTEST-BLAST>)

OliCheck has been tested and validated on Microsoft Windows™ XP. It may also run on Microsoft Windows™ 9x, Microsoft Windows™ NT, or Microsoft Windows™ 2000 (not tested).

The minimal OliCheck requirements are:

Pentium III processor (733MHz)  
128 MB RAM  
VGA or higher resolution monitor  
A mouse or other compatible pointing device

OliCheck is a memory and processor intensive application. As it uses multithreading for BLAST operations, OliCheck may also work with lower memory and processor requirements but this may yield poor performances.

The recommended configuration is the following:

Dual Xeon 1.5GHz processor  
1GB RAM  
1024x768 (32 bit) resolution monitor  
Windows™ XP SP1

The amount of RAM can be scaled according to the BLAST program requirements. The BLAST application loads the sequence databases in memory. When designing probes for larger genomes, the RAM size should be increased accordingly.

## Example Files

### ***BLAST Databases File***

The NCBI blast uses an internal database format that is generated from a simple FASTA file (See below) by using the `formatdb` tool. This tool generates a set of three files that are used by the `blastall` program as reference database. As an example, the command to generate the N315 database is the following:

```
formatdb -p F -n N315 -i N315.ffn
```

The parameter `-p F` indicate to `formatdb` that the input file contains nucleotide sequences. The `-n N315` parameters allows setting the output name of the database (i.e. the name that will be used in OliCheck BLAST Options). Finally the `-i N315.ffn` parameter setup the input file.

### ***FASTA File***

A FASTA file is a simple ASCII file. Every ORF entry is structured in the following way. The description line started by the character ">", it is followed by a free ASCII text. For OliCheck purposes, this text should be the accession name of the corresponding ORF. OliCheck uses this to annotate the output file if the *Add Hit Summary to Output* option is set. The following lines contain the sequence itself. The next ORF is recognized by the next > symbol.

A truncated example to a FASTA file is shown below:

```

>gi|15925706:517-1878, SA0001 [chromosomal replication initiator protein]
ATGTCGGAAGAAAGAAATTTGGGAAAAAGTCTTGAAATTTGCTCAAGAAAAATATCAGCTGTAAGTTACT
CAACTTTCTTAAAGACTACTGAGCTTTACACGATTAAAGATGGTGAAGCTATCGTATTATCGAGATTCC
TTTTAAATGCAAAATTTGGTTAAATCAACAAATATGCTGAAATTTATCCAAGCAATCTTATTGTGTTGTAGGC
TATGAAGTTAAACCTCACTTTTACTACTGAAGAATTAGCAAAATTATAGTAATATGAAGCTGTACTCT
CAAAAGAAACCAACAAACCTTTCTACTGAAACCACTGAGGATAATCATGTGCTTGGTAGAGAGCAATCAA
TGCCCATACACATTTGACACTTTTGTAAATCGGACCTGGTAACCGCTTTCCACATGCAGCAAGTTTAGCT
GTAGCCGAAGCACAGCCAAAGCGTACAATCCATTATTTATCTATGGAGGGTTGGTTTAGGAAAAACCC
ATTTAATGCATGCCATTGGTCATCATGTTTATAGATAATATCCAGATGCCAAAGTGATTTACACATCAAG
TGAAAAATTCACAAATGAATTTATTAAATCAATTCGTGATAACGAAGGTGAAGCTTTCAGAGAAAGATAT
CGTAATATCGACGCTCTTATTAAATCGATGATATTCAGTTTCATACAAAAATAAGTACAAACACAAGAAAGAT
TTTTCTATACCTTTTAAATGAATTCGATCAGAATAACAAGCAAAATAGTTATTTCGAGTGATCGACCAACAAA
GGAAATTCACAATTAGAAGACCGATTACGTTTCAACCTTTGAATGGGGCTAATTGTTGATATTACGCCA
CCAGATTATGAAATCCGAATGGCAATTTTCAGAGAAATTTGAAGAGAAATTTAGATATTCCACAG
AAGCTTTAAATTTATAGCAAAATCAAAATTCAAATTAATTTCTGTAATGAGAAGGTGCATTAACACGTTT
ACTTGCATATTCACAATTTATAGGAAAAACAAATTCACACTGAATTAACGTGCTGAAGCTTTAAAGATATTC
ATTCGAAGCAACAAATTCAAAAAGATTACCATCCAAAGATATTCAAAAAATTTGAGGCCAGTACTATAATG
TTAGAGATTGAAGATTTCAGTGCAAAAAACGTACAAGATCAATTCATATCCGCGTCAAAATAGCTATGTA
CTTGCTTAGAGAGCTTACAGATTCTCTATTACCTAAAAATTTGGTGAAGAATTTGGTGGCGGTGATCATACG
ACCGCTATTCATGCTCATGAAAAAATATCTAAAGATTAAAGAGATCCTATTTTAAACAAAGATAG
AGAAATCTTGAAAAAGAAATAAGAAATGTATAA
>gi|15925707:2156-3289, SA0002 [DNA polymerase III, beta chain]
ATGATGGAATTCACATTTAAAGAGATTATTTTATTACACAATTAATGACACATTAAGCTATTTCAC
CAAGAACCAACTTACCTATATTAACTGGTATCAAAATCGATGCGAAAGAACATGAAGTTATATACTACTGG
TTCAGACTCTGAAATTTCAATAGAAATCACTATTCTTAAACTGTAGATGGCGAAGATATTGTCAATATT
TCAGAAACAGGCTCAGTACTTCTCTGGACGATCTTTGTTGATATTATAAAAAAATTACCTGGTAAAG
ATGTTAAATTTATCTCAAAATGAACAAATTCAGACATTAATACATCAGGTCAATCTGAATTTAATTTAAG
TGGCTTAGATCCAGATCAATATCTTTATTACCTCAAGTTTCTAGAGATGACGCAATTCATTTGTGCGTA
AAAGTGCTTTAAAAACGTGATTGCAACAAACGAATTTTGCAGTGTCCACCTCAGAAACACGCCAGTACTAA
CTGGTGTGAACCTGGCTTATACAAGAAAAATGAATTAATATGCACAGCGACTGACTCACACCGCTTTGGCTGT
AAGAAAGTTGCAAGTTAGAGATGTTTCTGAAACAAAAATGTCAATTCAGGTAGGCTTTAGCTGAA
TTAAATTAATTTATGTTGCAATGAAGAGACATTTGATATCTTTTGGCTTCAACCAAGTTTATTATA
AAGTTGGAATGTGAACCTTTATTCTCGAATTTATGAAGGACATTTATCTGATACACACGCTTTATTCOC
TGAAACCTATGAATTAATTAAGTATAGACAATGGGAGTTTATCATGCGATTGATCGTGCTCTTTA
TTAGCAGGTGAAGGTGGTAAATACGTTTATTAATTAAGTACAGGTGATGACGTTGTTGAATTTATCTTCA
CATCACAGAAATTTGGTACTGTGAAAGAGAAAGTTGATGCAACGATGTTGAAGGTGGTAGCCTGAAAT
TTCAATTCACACTTAAATATATGATGGATGCTTTAAAGCAATCGATATGATGAGGTTGAAGTTGAATTC
TTCGGTCAATGAAACCAATTTATTCTAAAAACCAAGGTGACGACTCGGTACGCAATTAATTTTACCAA
TCAGAACTTACTAA
>gi|15925708:3760-3915, SA0003 [conserved hypothetical protein]
GTGATTATTTTGGTTCAAGAGTTGTAGTAGAAGGAGACATTAATTTAGGTCAATTTCTAAAAACAGAAG
GGATTATTGAATCTGGTGGTCAAGCAAAATGTTCTTGCAAGACGTTGAAGTATTAAATTAATGGAGTGGC
TGAAACACGTCGCGTAAAAAGTTAGAACATCAAGATCGTATAGATATCCAGAATTACCTGGAAGTGCT
GGTTCTTCTTAATCATTCATCAAGGTGAACAAATGA

```

## OliCheck Input File

OliCheck typical input file is an Array Designer™ output file (See description on page 8). Moreover, OliCheck can deal with any tab-delimited ASCII text file with the following restrictions. Each probe should be described in one line where columns 2 and 4 contain the accession and the sequence, respectively. The contents of the other columns can be freely filled as it is not considered as relevant by OliCheck. The output file generated by OliCheck can be directly set as an input file to the program, if the probes need to be further annotated.

## Array Designer™ Output File

Array Designer™ Output File is a 21 columns tab-delimited ASCII text file with the following form:

A truncated example of an Array Designed output file is shown below:

FileName	Accession	Site	Sense Probe	Rating	SeqPos	Length	Tm	GC%	Hairpin dG	SelfDimer dG	RunLen	--
NC_002745.fa	SA0280	100	TTGTAAAAGAATTGGAAAAGCATTAGTCAAAAGTATGATGAAATAGCTCAAAACTTTGGAA	44.3	50	60	69.63	28.3	-2.97	-2.97	4	--
NC_002745.fa	SA0280	100	TGTAAAAGAATTGGAAAAGCATTAGTCAAAAGTATGATGAAATAGCTCAAAACTTTGGAAA	44.3	51	60	69.63	28.3	-2.97	-2.97	4	--
NC_002745.fa	SA0280	100	GTAAGAAGATTGGAAAAGCATTAGTCAAAAGTATGATGAAATAGCTCAAAACTTTGGAAAA	43.8	52	60	69.2	28.3	-2.97	-2.97	4	--
NC_002745.fa	SA0474	50	ATGATTCAAGCATACTTAGGATTAGGTAGTAATATTGGTGATAGAGAAAGCCAGTTAAAC	60.2	1	60	69.79	33.3	-1.44	-2.62	3	--
NC_002745.fa	SA0474	50	ATTCAAGCATACTTAGGATTAGGTAGTAATATTGGTGATAGAGAAAGCCAGTTAAACGAT	60.1	4	60	70.32	33.3	-1.44	-2.62	3	--
NC_002745.fa	SA0668	100	TATTAAGACTCAAAAGAACTAACAGTTCAAGTTGTTGGTATTGATAATCTGCTGATAGA	59.8	54	60	70	31.7	-2.06	-2.06	3	--
NC_002745.fa	SA0668	100	ACTATATTAAAGACTCAAAAGAACTAACAGTTCAAGTTGTTGGTATTGATAATCTGCTGA	59.8	50	60	70.1	31.7	-2.06	-2.06	3	--
NC_002745.fa	SA0668	100	ATTAAGACTCAAAAGAACTAACAGTTCAAGTTGTTGGTATTGATAATCTGCTGATAGAA	59.7	55	60	70.29	31.7	-2.06	-2.06	3	--
NC_002745.fa	SA1414	200	CAGTTAAGTTAGTAGACAAAGCTGCTCAAAGTAATTTAATACATTCAAAACAAAGCTGACC	51.4	158	60	70.41	33.3	-2.93	-2.93	3	--
NC_002745.fa	SA1414	200	TAAGTTAGTAGACAAAGCTGCTCAAAGTAATTTAATACATTCAAAACAAAGCTGACCGTAT	51.3	162	60	70.49	31.7	-2.93	-2.93	3	--
NC_002745.fa	SA1414	200	TTAAGTTAGTAGACAAAGCTGCTCAAAGTAATTTAATACATTCAAAACAAAGCTGACCGTA	51.3	161	60	70.55	31.7	-2.93	-2.93	3	--
NC_002745.fa	SA1220	150	GCGATTCCATTTTAAACAACACTAGGTATATTAATCACATTGCTTTTAGAAACCATCACT	51.7	103	60	70.24	31.7	-1.61	-2.62	4	--
NC_002745.fa	SA1220	150	TCAGCGATTCCATTTTAAACAACACTAGGTATATTAATCACATTGCTTTTAGAAACCATC	51.4	100	60	70.14	31.7	-1.7	-2.62	4	--
NC_002745.fa	SA1608	350	TGGATAAAGCATTAGAGTATCGTGATAAGATAGCGAAGAAGAAATTGAAGCAACAGGTG	57.6	326	60	72.01	36.7	-1.71	-1.71	3	--
Antisense Probe				Rating	SeqPos	Length	Tm	GC%	Hairpin dG	SelfDimer dG	RunLen	
TTCCAAAGTTTTGAGCTATTTTCATCATACTTTTGACTAATGCTTTCCAATTCCTTTACAA				44.3	109	60	69.63	28.3	-2.97	-2.97	4	
TTTCCAAAGTTTTGAGCTATTTTCATCATACTTTTGACTAATGCTTTCCAATTCCTTTACA				44.3	110	60	69.63	28.3	-2.97	-2.97	4	
TTTTCCAAAGTTTTGAGCTATTTTCATCATACTTTTGACTAATGCTTTCCAATTCCTTTTAC				43.8	111	60	69.2	28.3	-2.97	-2.97	4	
GTTTAACTGGCTTTCTCTATCACCAATATTACTACCTAATCCTAAGTATGCTTGAATCAT				60.2	60	60	69.79	33.3	-1.44	-2.62	3	
ATCGTTTAACTGGCTTTCTCTATCACCAATATTACTACCTAATCCTAAGTATGCTTGAAT				60.1	63	60	70.32	33.3	-1.44	-2.62	3	
TCTATCAGCAGATTATCAATACCAACAACCTTGAACCTGTTAGTTTCGTTTGAGTCTTAATA				59.8	113	60	70	31.7	-2.06	-2.06	3	
TCAGCAGATTATCAATACCAACAACCTTGAACCTGTTAGTTTCGTTTGAGTCTTAATATAGT				59.8	109	60	70.1	31.7	-2.06	-2.06	3	
TTCTATCAGCAGATTATCAATACCAACAACCTTGAACCTGTTAGTTTCGTTTGAGTCTTAAT				59.7	114	60	70.29	31.7	-2.06	-2.06	3	
GGTCAGCTTTGTTTGAATGTATTAAATTACTTTGAGCAGCTTTGTCTACTAACTTAACTG				51.4	217	60	70.41	33.3	-2.93	-2.93	3	
ATACGGTCAGCTTTGTTTGAATGTATTAAATTACTTTGAGCAGCTTTGTCTACTAACTTA				51.3	221	60	70.49	31.7	-2.93	-2.93	3	
TACGGTCAGCTTTGTTTGAATGTATTAAATTACTTTGAGCAGCTTTGTCTACTAACTTAA				51.3	220	60	70.55	31.7	-2.93	-2.93	3	
AGTGATGGTTTCTAAAAGCAATGTGATTAAATACCTAGTGTGTTTAAAATGGAAATCGC				51.7	162	60	70.24	31.7	-1.61	-2.62	4	
GATGGTTTCTAAAAGCAATGTGATTAAATACCTAGTGTGTTTAAAATGGAAATCGCTGA				51.4	159	60	70.14	31.7	-1.7	-2.62	4	
CACCTGTGTGCTTCAATTTCTCTTCGCTATCTTTATCAGCATACTCTAATGCTTTATCCA				57.6	385	60	72.01	36.7	-1.71	-1.71	3	

## OliCheck Passed Output Files

OliCheck output file format is derived from the input file. OliCheck simply copies the line corresponding to the probe in the output file. If the *Add Hit Summary to Output* option is set, all input lines are copied but only the Probes that passed OliCheck filters are annotated. If the probe fails at any stage during the process, the annotation is left blank. Alternatively

if the *Add Hit Summary to Output* option is unchecked, the probes that failed at any stage during the OliCheck process are discarded from the Passed output file.

## Annotated Passed File

Annotated passed file adds a new column showing the accession name of the corresponding homologous hit.

A truncated example of an OliCheck annotated output file is shown below:

FileName	Accession	Site	Sense Probe	Rating	SeqPos	Length	Tm	GC%	Hairpin dG	SelfDimer dG	RunLen	--
NC_002745.fa	SA0280	100	TTGTAAAAGAATTGGAAAGCATTAGTCAAAAGTATGATGAAATAGCTCAAAACTTTTGAA	44.3	50	60	69.63	28.3	-2.97	-2.97	4	--
NC_002745.fa	SA0280	100	TGTAAAAGAATTGGAAAGCATTAGTCAAAAGTATGATGAAATAGCTCAAAACTTTTGAA	44.3	51	60	69.63	28.3	-2.97	-2.97	4	--
NC_002745.fa	SA0280	100	GTAAAAGAATTGGAAAGCATTAGTCAAAAGTATGATGAAATAGCTCAAAACTTTTGAA	43.8	52	60	69.2	28.3	-2.97	-2.97	4	--
NC_002745.fa	SA0474	50	ATGATTCAAGCATACTTAGGATTAGTGTAGTAATATTGGTGATAGAGAAAGCCAGTTAAAC	60.2	1	60	69.79	33.3	-1.44	-2.62	3	--
NC_002745.fa	SA0474	50	ATTCAGCATACTTAGGATTAGTGTAGTAATATTGGTGATAGAGAAAGCCAGTTAAACGAT	60.1	4	60	70.32	33.3	-1.44	-2.62	3	--
NC_002745.fa	SA0668	100	TATTAAGACTCAAAAGAACTAACAGTTCAAGTTGTTGGTATTGATTAATCTGCTGATAGA	59.8	54	60	70	31.7	-2.06	-2.06	3	--
NC_002745.fa	SA0668	100	ACTATATTAAAGACTCAAAAGAACTAACAGTTCAAGTTGTTGGTATTGATTAATCTGCTGA	59.8	50	60	70.1	31.7	-2.06	-2.06	3	--
NC_002745.fa	SA0668	100	ATTAAGACTCAAAAGAACTAACAGTTCAAGTTGTTGGTATTGATTAATCTGCTGATAGA	59.7	55	60	70.29	31.7	-2.06	-2.06	3	--
NC_002745.fa	SA1414	200	CAGTTAAGTTAGTAGACAAGCTGCTCAAGTAAATTTAATACATTCAAAAGCTGACCC	51.4	158	60	70.41	33.3	-2.93	-2.93	3	--
NC_002745.fa	SA1414	200	TAAGTTAGTAGACAAGCTGCTCAAGTAAATTTAATACATTCAAAAGCTGACCCGAT	51.3	162	60	70.49	31.7	-2.93	-2.93	3	--
NC_002745.fa	SA1414	200	TTAAGTTAGTAGACAAGCTGCTCAAGTAAATTTAATACATTCAAAAGCTGACCCGTA	51.3	161	60	70.55	31.7	-2.93	-2.93	3	--
NC_002745.fa	SA1220	150	GCGATTTCCATTTTAAACAACACTAGGTATATTAATCACATTGCTTTTAGAAACCATCACT	51.7	103	60	70.24	31.7	-1.61	-2.62	4	--
NC_002745.fa	SA1220	150	TCAGCGATTTCCATTTTAAACAACACTAGGTATATTAATCACATTGCTTTTAGAAACCATC	51.4	100	60	70.14	31.7	-1.7	-2.62	4	--
NC_002745.fa	SA1608	350	TGGATAAAGCATTAGAGTATCGTGATAAAGATAGCGAAGAAGAAATTGAAGCAACAGTG	57.6	326	60	72.01	36.7	-1.71	-1.71	3	--
NC_002745.fa	SA1608	350	GTGGATAAAGCATTAGAGTATCGTGATAAAGATAGCGAAGAAGAAATTGAAGCAACAGGT	57.6	325	60	72.01	36.7	-1.71	-1.71	3	--
NC_002745.fa	SA1608	350	CACAAGCGGTGGATAAAGCATTAGAGTATCGTGATAAAGATAGCGAAGAAGAAATTGAAG	56.3	317	60	72.27	38.3	-1.71	-1.71	3	--
NC_002745.fa	SA1608	350	CGTGATAAAGCATTAGAGTATCGTGATAAAGATAGCGAAGAAGAAATTGAAGCAACAGG	56.3	324	60	72.27	38.3	-1.71	-1.71	3	--

--	Antisense Probe	Rating	SeqPos	Length	Tm	GC%	Hairpin dG	SelfDimer dG	RunLen	
--	TTCCAAAGTTTGTAGCTATTTTCATCATACTTTTGACTAATGCTTTCCAAATTCCTTTACAA	44.3	109	60	69.63	28.3	-2.97	-2.97	4	SA0499
--	TTTCCAAAGTTTGTAGCTATTTTCATCATACTTTTGACTAATGCTTTCCAAATTCCTTTTACA	44.3	110	60	69.63	28.3	-2.97	-2.97	4	SA1225
--	TTTTCCAAAGTTTGTAGCTATTTTCATCATACTTTTGACTAATGCTTTCCAAATTCCTTTTAC	43.8	111	60	69.2	28.3	-2.97	-2.97	4	SA1221
--	GTTTAACTGGCTTTCTCTATCACCAATATTACTACCTAATCCTAAGTATGCTTGAATCAT	60.2	60	60	69.79	33.3	-1.44	-2.62	3	SA1439
--	ATCGTTTAACTGGCTTTCTCTATCACCAATATTACTACCTAATCCTAAGTATGCTTGAAT	60.1	63	60	70.32	33.3	-1.44	-2.62	3	SA1438
--	TCTATCAGCAGATTATCAATACCAACAACCTTGAACGTGTAGTTTCGTTTGAGTCTTAATA	59.8	113	60	70	31.7	-2.06	-2.06	3	SA1433
--	TCAGCAGATTATCAATACCAACAACCTTGAACGTGTAGTTTCGTTTGAGTCTTAATATAGT	59.8	109	60	70.1	31.7	-2.06	-2.06	3	SA1427
--	TTCTATCAGCAGATTATCAATACCAACAACCTTGAACGTGTAGTTTCGTTTGAGTCTTAAT	59.7	114	60	70.29	31.7	-2.06	-2.06	3	SA1420
--	GGTCAGCTTTGTTTGAATGTATTAATTAATTTTGAAGCAGCTTTTGCTACTAATCTTAACGTG	51.4	217	60	70.41	33.3	-2.93	-2.93	3	
--	ATACGGTCAGCTTTGTTTGAATGTATTAATTAATTTTGAAGCAGCTTTTGCTACTAATCTTA	51.3	221	60	70.49	31.7	-2.93	-2.93	3	
--	TACGGTCAGCTTTGTTTGAATGTATTAATTAATTTTGAAGCAGCTTTTGCTACTAATCTTA	51.3	220	60	70.55	31.7	-2.93	-2.93	3	SA0883
--	AGTGATGGTTTCTAAAAGCAATGTGATTAATATACCTAGTGTGTTAAAATGGAAATCGC	51.7	162	60	70.24	31.7	-1.61	-2.62	4	SA0871
--	GATGGTTTCTAAAAGCAATGTGATTAATATACCTAGTGTGTTAAAATGGAAATCGCTGA	51.4	159	60	70.14	31.7	-1.7	-2.62	4	SA0869
--	CACCTGTGCTTCAATTTCTTCTCGCTATCTTTATCAGGATACCTTAATGCTTTATCCCA	57.6	385	60	72.01	36.7	-1.71	-1.71	3	
--	ACCTGTGCTTCAATTTCTTCTCGCTATCTTTATCAGGATACCTTAATGCTTTATCCCA	57.6	384	60	72.01	36.7	-1.71	-1.71	3	SA0866
--	CTTCAATTTCTTCTCGCTATCTTTATCAGGATACCTTAATGCTTTATCCACGCTTGTG	56.3	376	60	72.27	38.3	-1.71	-1.71	3	SA1611

## Simple Passed File

Simple passed file removes the probes that failed OliCheck filters from the input file.

A truncated example of an OliCheck output file is shown below:

FileName	Accession	Site	Sense Probe	Rating	SeqPos	Length	Tm	GC%	Hairpin dG	SelfDimer dG	RunLen	--
NC_002745.fa	SA0280	100	TTGTAAAAGAATTGGAAAGCATTAGTCAAAAGTATGATGAATAGCTCAAAACTTTGGAA	44.3	50	60	69.63	28.3	-2.97	-2.97	4	--
NC_002745.fa	SA0280	100	TGTAAAAGAATTGGAAAGCATTAGTCAAAAGTATGATGAATAGCTCAAAACTTTGGAA	44.3	51	60	69.63	28.3	-2.97	-2.97	4	--
NC_002745.fa	SA0280	100	GTAAAAGAATTGGAAAGCATTAGTCAAAAGTATGATGAATAGCTCAAAACTTTGGAA	43.8	52	60	69.2	28.3	-2.97	-2.97	4	--
NC_002745.fa	SA0474	50	ATGATTCAAGCATACTTAGGATTAGGTAGTAATATTGGTGATAGAGAAAGCCAGTTAAAC	60.2	1	60	69.79	33.3	-1.44	-2.62	3	--
NC_002745.fa	SA0474	50	ATTCAAGCATACTTAGGATTAGGTAGTAATATTGGTGATAGAGAAAGCCAGTTAAACGAT	60.1	4	60	70.32	33.3	-1.44	-2.62	3	--
NC_002745.fa	SA0668	100	TATTAAGACTCAAAACGAACTAACAGTTCAAGTTGTTGGTATTGATAATCTGCTGATAGA	59.8	54	60	70	31.7	-2.06	-2.06	3	--
NC_002745.fa	SA0668	100	ACTATATTAAAGACTCAAAACGAACTAACAGTTCAAGTTGTTGGTATTGATAATCTGCTGA	59.8	50	60	70.1	31.7	-2.06	-2.06	3	--
NC_002745.fa	SA0668	100	ATTAAGACTCAAAACGAACTAACAGTTCAAGTTGTTGGTATTGATAATCTGCTGATAGAA	59.7	55	60	70.29	31.7	-2.06	-2.06	3	--
NC_002745.fa	SA1414	200	TTAAGTTAGTAGACAAAGCTGCTCAAAGTAATTTAATACATTCAAAACAAAGCTGACCGTA	51.3	161	60	70.55	31.7	-2.93	-2.93	3	--
NC_002745.fa	SA1220	150	GCGATTCCATTTTAACAACACTAGGTATATTAATCACATTGCTTTTAGAAACCATCACT	51.7	103	60	70.24	31.7	-1.61	-2.62	4	--
NC_002745.fa	SA1220	150	TCAGCGATTTCATTTTAAACAACACTAGGTATATTAATCACATTGCTTTTAGAAACCATC	51.4	100	60	70.14	31.7	-1.7	-2.62	4	--
NC_002745.fa	SA1608	350	GTGGATAAAGCATTAGAGTATCGTGATAAAGATAGCGAAGAAGAAATTTGAAGCAACAGGT	57.6	325	60	72.01	36.7	-1.71	-1.71	3	--
NC_002745.fa	SA1608	350	CACAAGCGGTGGATAAAGCATTAGAGTATCGTGATAAAGATAGCGAAGAAGAAATTTGAAG	56.3	317	60	72.27	38.3	-1.71	-1.71	3	--
NC_002745.fa	SA1608	350	CGTGGATAAAGCATTAGAGTATCGTGATAAAGATAGCGAAGAAGAAATTTGAAGCAACAGG	56.3	324	60	72.27	38.3	-1.71	-1.71	3	--
...												
Antisense Probe				Rating	SeqPos	Length	Tm	GC%	Hairpin dG	SelfDimer dG	RunLen	
TTCCAAAGTTTGTAGCTATTTTCATCATACTTTTGACTAATGCTTTCCAATTCTTTTACAA				44.3	109	60	69.63	28.3	-2.97	-2.97	4	
TTTCCAAAGTTTGTAGCTATTTTCATCATACTTTTGACTAATGCTTTCCAATTCTTTTACA				44.3	110	60	69.63	28.3	-2.97	-2.97	4	
TTTTCCAAAGTTTGTAGCTATTTTCATCATACTTTTGACTAATGCTTTCCAATTCTTTTAC				43.8	111	60	69.2	28.3	-2.97	-2.97	4	
GTTTAACTGGCTTCTCTATCACCAATATTACTACCTAATCCTAAGTATGCTTGAATCAT				60.2	60	60	69.79	33.3	-1.44	-2.62	3	
ATCGTTTAACTGGCTTCTCTATCACCAATATTACTACCTAATCCTAAGTATGCTTGAAT				60.1	63	60	70.32	33.3	-1.44	-2.62	3	
TCTATCAGCAGATTATCAATACCAACAACCTTGAACCTGTTAGTTTCGTTTGAGTCTTAATA				59.8	113	60	70	31.7	-2.06	-2.06	3	
TCAGCAGATTATCAATACCAACAACCTTGAACCTGTTAGTTTCGTTTGAGTCTTAATATAGT				59.8	109	60	70.1	31.7	-2.06	-2.06	3	
TTCTATCAGCAGATTATCAATACCAACAACCTTGAACCTGTTAGTTTCGTTTGAGTCTTAAT				59.7	114	60	70.29	31.7	-2.06	-2.06	3	
TACGCTCAGCTTGTGTAATGTATTAATTAATCTTGAGCAGCTTTGTCTACTAACTTAA				51.3	220	60	70.55	31.7	-2.93	-2.93	3	
AGTGATGGTTTCTAAAAGCAATGTGATTAATATACCTAGTGTGTTTAAAATGGAAATCGC				51.7	162	60	70.24	31.7	-1.61	-2.62	4	
GATGGTTTCTAAAAGCAATGTGATTAATATACCTAGTGTGTTTAAAATGGAAATCGCTGA				51.4	159	60	70.14	31.7	-1.7	-2.62	4	
ACCTGTGCTTCAATTCTTCTTCGCTATCTTTATCACGATACTCTAATGCTTTATCCAC				57.6	384	60	72.01	36.7	-1.71	-1.71	3	
CTTCAATTTCTTCTTCGCTATCTTTATCACGATACTCTAATGCTTTATCCAGCCTTGTG				56.3	376	60	72.27	38.3	-1.71	-1.71	3	
CCTGTGCTTCAATTTCTTCTTCGCTATCTTTATCACGATACTCTAATGCTTTATCCACG				56.3	383	60	72.27	38.3	-1.71	-1.71	3	

## OliCheck Failed Output Files

The failed output file is a list of accession numbers for which no probes have been found valid according to OliCheck filter parameters. This file is not generated if the *Add Hit Summary to Output* option is checked.

A truncated example of an OliCheck failed output file is shown below:

```
SA1248  
SA1245  
SA0498  
SA0496  
SA0494  
SA0493  
SA1240  
SA0489  
SA1236  
SA1235  
SA0487  
SA1234  
SA0486  
SA0485  
SA1233  
SA0484  
SA1231
```